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Calcium montmorillonite clay reduces urinary biomarkers of fumonisin B₁ exposure in rats and humans

A. Robinson^a, N.M. Johnson^{a†}, A. Strey^b, J.F. Taylor^a, A. Marroquin-Cardona^a, N.J. Mitchell^a, E. Afriyie-Gyawu^c, N.A. Ankrah^d, J.H. Williams^e, J.S. Wang^f, P.E. Jolly^g, R.J. Nachman^b and T.D. Phillips^{a*}

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Fumonisin B₁ (FB₁) is often a co-contaminant with aflatoxin (AF) in grains and may enhance AF's carcinogenicity by acting as a cancer promoter. Calcium montmorillonite (i.e. NovaSil, NS) is a possible dietary intervention to help decrease chronic aflatoxin exposure where populations are at risk. Previous studies show that an oral dose of NS clay was able to reduce AF exposure in a Ghanaian population. *In vitro* analyses from our laboratory indicated that FB₁ (like aflatoxin) could also be sorbed onto the surfaces of NS. Hence, our objectives were to evaluate the efficacy of NS clay to reduce urinary FB₁ in a rodent model and then in a human population highly exposed to AF. In the rodent model, male Fisher rats were randomly assigned to either FB₁ control, FB₁ + 2% NS or absolute control group. FB₁ alone or with clay was given as a single dose by gavage. For the human trial, participants received NS (1.5 or 3 g day⁻¹) or placebo (1.5 g day⁻¹) for 3 months. Urines from weeks 8 and 10 were collected from the study participants for analysis. In rats, NS significantly reduced urinary FB₁ biomarker by 20% in 24 h and 50% after 48 h compared to controls. In the humans, 56% of the urine samples analysed (*n* = 186) had detectable levels of FB₁. Median urinary FB₁ levels were significantly (*p* < 0.05) decreased by >90% in the high dose NS group (3 g day⁻¹) compared to the placebo. This work indicates that our study participants in Ghana were exposed to FB₁ (in addition to AFs) from the diet. Moreover, earlier studies have shown conclusively that NS reduces the bioavailability of AF and the findings from this study suggest that NS clay also reduces the bioavailability FB₁. This is important since AF is a proven dietary risk factor for hepatocellular carcinoma (HCC) in humans and FB₁ is suspected to be a dietary risk factor for HCC and oesophageal cancer in humans.

Keywords: HPLC; toxicology, animal study; clinical study; aflatoxins; fumonisins; mycotoxins

Introduction

Fumonisin B₁ (FB₁) is the most abundant of the naturally occurring fumonisins. FB₁ is produced by *Fusarium* fungi and it has been shown to be hepatotoxic, nephrotoxic and carcinogenic in a number of species (Voss et al. 2002). Epidemiological studies have correlated *Fusarium* spp. and fumonisin contamination of food sources with increased incidences of oesophageal cancer in regions of China and South Africa, neural tube defects along the Texas–Mexico border, and primary liver cancer in China (Chu and Li 1994; Ueno et al. 1997; Marasas et al. 2004; Shephard et al. 2007). However, there are no reports that definitively demonstrate a causative relationship (Stockmann-Juvala and Savolainen 2008). While methods assessing

the hazard of fumonisins in foodstuffs exist, few are capable of determining the actual exposure of populations considered to be “at risk” (Shephard et al. 1996, 2007). Changes in sphingolipid ratio due to the inhibitory effect of FB₁ on *de novo* ceramide synthase activity are commonly utilised as biomarkers for FB₁ exposure (Voss et al. 2002; He et al. 2006; Sabourdy et al. 2008). Alterations in the sphinganine:sphingosine ratio have accurately reflected fumonisin exposure in laboratory and farm animals, but have not been shown to be consistent indicators of exposure in human populations consuming fumonisin-contaminated foods (Abnet et al. 2001; Solfrizzo et al. 2004). Metabolic studies in non-human primates and swine have shown that excretion of FB₁ is mainly through the faeces with ≤1% urinary excretion (Shephard et al. 1994;

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Fodor et al. 2008). Due to high dietary levels of FB₁ in developing countries, urinary biomarkers have been successfully used to characterise exposure in human populations (Shetty et al. 1998; Turner et al. 1999; Gong et al. 2008; Van der Westhuizen et al. 2011). This biomarker has been applied to evaluate intervention strategies that could reduce exposure to fumonisins (Van der Westhuizen et al. 2011). Our laboratory has previously reported that a Ghanaian population is highly exposed to aflatoxins (AFs) due to the frequent consumption of AF-contaminated foods (Jolly et al. 2006; Phillips et al. 2008; Wang et al. 2008). It has been well-documented that AFs are contributors of immunosuppression, malnutrition and hepatocellular carcinoma (Wogan 1992; Williams et al. 2004; Jiang et al. 2005). Furthermore, *in vitro* and *in vivo* studies have demonstrated that FB₁ can potentiate the effects of AFs (Carlson et al. 2001; IARC 2002; McKean et al. 2006). Kpodo et al. (2000) verified the co-occurrence of *Fusarium* spp. and fumonisins with AFs in maize samples from Ghanaian markets. Hence, it was postulated that participants from our previous study in Ghana, shown to be at high risk for aflatoxicosis, may be co-exposed to fumonisins. To reduce AF exposure, the use of NovaSil (NS) (a dioctahedral smectite clay) as an intervention plan for the enterosorption of the toxin has been shown to be safe and effective in humans (Phillips et al. 2008). Importantly, NS has also been evaluated *in vitro* for FB₁ sorption and found to interact with this mycotoxin at interlayer surfaces at acidic pH (Lemke 2000). A possible mechanism for FB₁ sorption to NS is protonation of the amino group at C2 on the molecule in acidic conditions (i.e. the stomach). The charged FB₁ may be bound through a cationic exchange reaction at negatively charged surfaces of the clay.

Thus, the main objectives of this study were to: (1) determine the efficacy of NS to reduce urinary FB₁ in Fischer 344 rats; (2) assess exposure to fumonisins in a Ghanaian population highly exposed to AFs using the FB₁ urinary biomarker; and (3) assess the efficacy of NS to reduce fumonisin exposure in this population. Importantly, strategies that reduce co-exposures to AFs and fumonisins are highly desirable for populations at risk for both agents.

Materials and methods

Chemicals

FB₁, *o*-phthalaldehyde (OPA), methanol, 2-mercaptoethanol, sodium phosphate dibasic and phosphoric acid were purchased from Sigma-Aldrich Chemical Co. (St. Louis, MO, USA). Naphthalene 2,3-dicarboxaldehyde (NDA), potassium cyanide (KCN), sodium tetraborate (STB), phosphate-buffered saline (PBS) and potassium hydroxide (KOH) were also obtained

from Sigma. Fumonitest WB columns were purchased from VICAM (Watertown, MA, USA). All of the experiments were done using filtered and deionised water (18.2 MΩ·cm) (Millipore, Milford, MA, USA), and all other chemicals and reagents were purchased commercially at the highest degree of purity available.

Preparation of FB₁ standard and derivatisation solutions

FB₁ standard solution (1 mg ml⁻¹) was prepared in 50:50 (v/v) methanol/water. NDA and OPA solutions were made daily and stored at 4 °C in the dark. NDA was prepared as described by He et al. (2005) and OPA was prepared according to Cai et al. (2007). These solutions were prepared according to the above mentioned methods, except during the fluorescence comparison analysis in which equal molar concentrations of NDA and OPA were used. For the experiments comparing sensitivity of the derivatising agents, excitation and emission parameters were set at 335 and 440 nm for OPA and 252 and 483 nm for NDA.

Rodent model experimental design

A total of 18 male Fisher 344 rats (5-week-old male, 110–130 g body weight, bw) were purchased from Harlan (Houston, TX, USA) and maintained on nutritionally complete powdered feed (Teklad rodent diet 8604; Harlan, Madison, WI, USA) and water *ad libitum*. The rats were randomised into three equal groups: (A) Absolute control; (B) FB₁ control; and (C) FB₁ plus 2% (w/w) NS inclusion in the feed. The dietary NS clay concentration was based on the highest level previously determined to be safe in a chronic rodent study (Afriyie-Gyawu et al. 2005). After a brief acclimation period (1 week), the rats were gavaged and placed in metabolism cages (1 rat per cage) for 8 days and daily urine samples were collected. Groups B and C were administered 3.75 mg FB₁ per rat (25 mg kg⁻¹ bw based on an average of 150 g bw) via aqueous gavage. The gavage solution for group C also contained 2% NS (the daily amount inclusion based on an average of 20 g feed consumed per day). Group C also received dietary NS (2% wet weight, ww) during the acclimation week and the week after gavage. All rats were housed in a climate-controlled environment (temperature 22–25 °C) that was artificially illuminated (12-h dark/light cycle) and free from chemical contamination. Rats were inspected daily for general appearance, behavioural changes and signs of morbidity and mortality. Body weights were measured prior to acclimation, prior to gavage, and every 2 days during sample collection.

Human study site and population

Stored urine samples were available from our previous Phase IIa clinical trial assessing the safety and efficacy of NovaSil for reducing AF exposure in a Ghanaian population (Afriyie-Gyawu et al. 2008; Phillips et al. 2008). Volunteers were recruited from six communities within the Ejura-Sekyedumase district in the Ashanti Region of Ghana. Biomarker data regarding exposure to fumonisins was unknown for this area; however, maize samples from neighbouring districts in Ghana had been shown to contain fumonisins (Kpodo et al. 2000). To our knowledge, this is the first study to measure FB₁ in a human population in Ghana. A total of 177 volunteers (male and female) were selected to participate in the NS intervention study based on predetermined inclusion criteria. Participants met the following conditions: signed consent form, healthy status based on physical examination, age 18–58 years, intake of corn and/or groundnut-based foods at least four times per week, blood AFB₁-albumin adduct levels >0.5 pmol AFB₁ mg⁻¹ albumin, no history of chronic disease(s), no use of prescribed medications, non-pregnant and non-breastfeeding females, normal ranges of haematological parameters and normal liver and renal function indicators (blood and urine parameters). The FB₁ levels in food consumed by our study participants were not measured and hence could not be correlated with urinary excretion patterns.

Human study design and protocol

The study protocol was approved by the Institutional Review Boards at Texas A&M University and Noguchi Memorial Institute for Medical Research in Ghana for Ethical Clearance. All participants were provided written informed consent, as well as an oral explanation prior to beginning the study. The overall study design adhered to guidelines set for a randomised, double-blind, placebo controlled Phase IIa clinical trial (Sangare-Tigori et al. 2006). Participants were randomly divided into one of three groups: High dose (NS 3.0 g day⁻¹), low dose (NS 1.5 g day⁻¹) or placebo control (1.5 g day⁻¹; microcrystalline cellulose) for a period of 3 months. Trained study monitors collected blood and urine samples (morning urine) from each participant at multiple time points. Aliquots of the urines were stored separately in polypropylene tubes and shipped frozen to Texas A&M University where they were stored at -80°C prior to biomarker analysis. Samples collected at weeks 8 and 10 from all three treatment groups (186 samples) were used in this preliminary study to assess fumonisin exposure and intervention efficacy. Baseline urine samples were not available for analysis.

FB₁ urine analysis

Frozen urine samples were thawed at room temperature and centrifuged at 500 g for 5 min. Portions (10 ml) of the urine samples were passed through Fumonitest WB columns (VICAM, Watertown, MA, USA) at a flow rate of approximately 1 ml min⁻¹. Columns were washed with 6 ml phosphate-buffered saline and 6 ml of water before elution with 2 ml of 100% methanol. Samples were dried at 50°C under nitrogen gas and reconstituted in 150 µl of 50:50 (v/v) methanol/water. Samples were derivatised with NDA as previously described by Cho et al. (2002). Briefly, aliquots of NDA solution (300 µl) were added to samples which were vortexed and heated (50°C) for 10 min. Analyte extracts were injected (200 µl) onto a 250 × 4.6 mm C₁₈ LUNA column, 5 µm particle size (Phenomenex, Torrance, CA, USA) with mobile phase consisting of 77:23 (v/v) methanol/sodium phosphate dibasic (0.1 M, pH 3.35) at a flow-rate of 1 ml min⁻¹. FB₁ analysis was performed using a Waters HPLC System equipped with an auto-injector (model 717A) and fluorescence detector (model 2475). Excitation and emission parameters were set at 252 and 483 nm for detection of NDA-derivatised samples. Creatinine concentrations in animal urine samples were measured at the Texas Veterinary Medical Diagnostic Laboratory in College Station (TX, USA). Creatinine concentrations in human urine samples were measured at St. Joseph's Regional Health Center Laboratory in Bryan (TX, USA). The identity of the urine metabolite was confirmed via MALDI-TOF on a Kratos Kompact Probe MALDI-MS instrument (Kratos Analytical). A saturated solution of alpha-cyano-4-hydroxycinnamic acid (in methanol) was used to facilitate ionisation of the sample as previously described by Neupert et al. (2009).

Statistics

Data generated from HPLC analyses were transferred into an Excel database for management. Mean, median, confidence interval (CI) and range were calculated for concentrations of FB₁. To show the effect of NS ingestion on FB₁ levels, statistical evaluation focused on the comparisons among different treatment groups in the rodent model and human population. Two time points were utilised to further evaluate the efficacy of the NS sorbent in the human clinical intervention trial. For statistical analysis, non-detectable samples = LOD/2. The Kruskal-Wallis and Wilcoxon rank-sum non-parametric tests were used to compare the differences among and between treatment groups. A *p*-value ≤0.05 (two-tailed) was considered significant. Statistical analyses were done using SPSS software version 15.0 (SPSS Inc., Chicago, IL, USA).

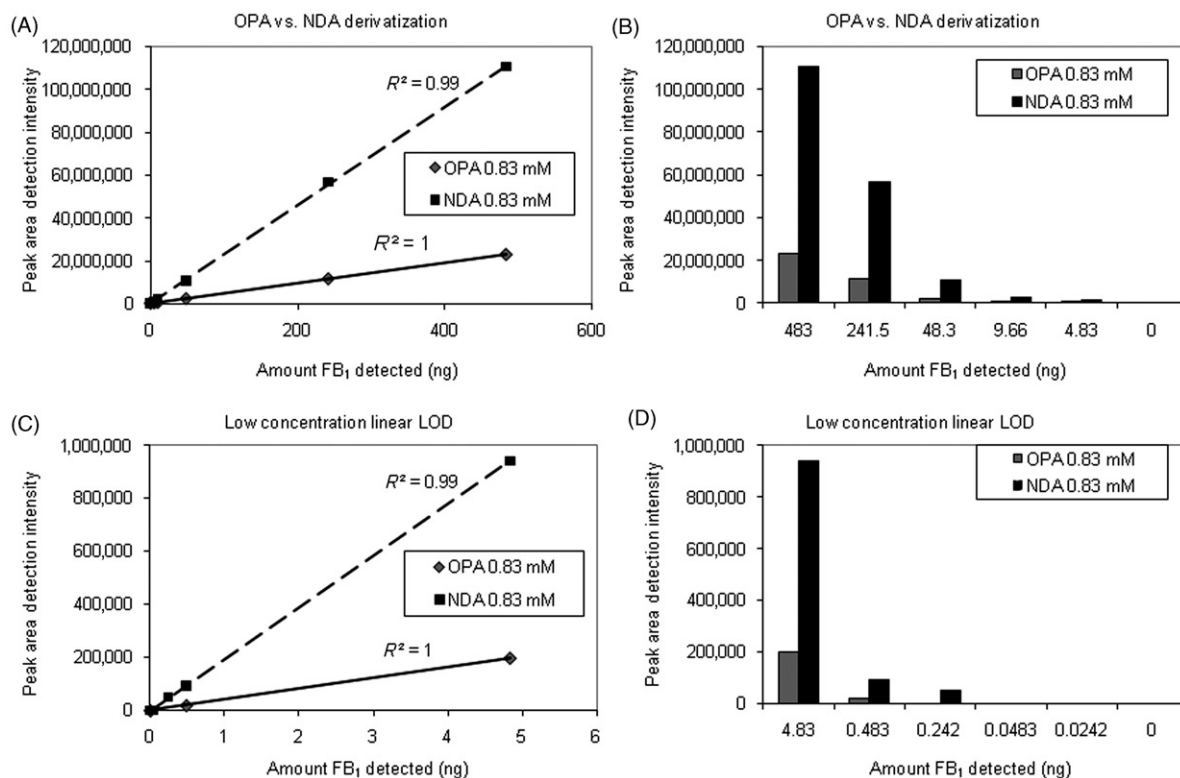


Figure 1. Comparison of FB₁ with OPA derivatisation vs. NDA derivatisation. Both OPA and NDA derivatisation products were linear within a range of 483–4.83 ng (A and B). The relative intensity of NDA derivatisation was 2–5 times higher than that of OPA (concentration dependent) allowing for a lower limit of detection (0.242 ng) while retaining linearity (C and D).

Results

Comparison of OPA versus NDA derivatisation sensitivity

Detection of both OPA and NDA derivatised FB₁ was linear (0–480 ng), with NDA having a stable lower LOD (0.242 ng) (Figure 1A–D). The detected relative intensity of NDA derivatised FB₁ was two to five times the intensity of OPA derivatised FB₁ (Figure 1A, B). Thus, NDA derivatisation was used and favoured for analysis of urine samples in both the rodent model and human study.

NovaSil efficacy in the rodent model

Urine collected at 24, 48 and 72 h post-gavage were analysed for FB₁. Most of the FB₁ dosed was eliminated in urine within 72 h (Figure 2). At 24 h, the mean total urinary FB₁ excretion for rats in group B was 864.7 ng FB₁ ml⁻¹ urine (194.1 ng FB₁ mg⁻¹ creatinine). After 48 h, excretion of the biomarker was reduced to 183.1 ng FB₁ ml⁻¹ urine (9.4 ng FB₁ mg⁻¹ creatinine). Rats in group C showed a mean total urinary excretion of 625.7 ng FB₁ ml⁻¹ urine (158.2 ng FB₁ mg⁻¹ creatinine) at 24 h post-gavage. After 48 h, the FB₁ excretion was reduced to 83.3 ng FB₁ ml⁻¹ urine (4.4 ng FB₁ mg⁻¹ creatinine). The urines from

animals in group A were used to verify the lack of urinary FB₁ (Table 1). There was a statistically significant difference ($p < 0.05$) in the median (Mdn) levels of urinary FB₁ when all three treatment groups were compared. At 24 h post-gavage, median value of urinary FB₁ was significantly reduced by 34% in the NS treatment group (Mdn = 590.4 ng ml⁻¹ urine) compared to the FB₁ control group (Mdn = 892.4 ng ml⁻¹ urine) ($p \leq 0.05$). When standardised with creatinine, urinary FB₁ was still significantly reduced by 20%. At 48 h post-gavage, there was a significant reduction of 50% in urinary FB₁ in the NS treatment group when compared to the FB₁ control group.

NovaSil efficacy in the human study

A total of 186 urine samples from study participants at week 8 and week 10 were analysed for the presence of FB₁. Mean and median levels of FB₁ are presented in Table 2. When placebo groups were compared, 60.9% of week 8 (14/23) and 72.1% of week 10 (31/43), samples were found to be positive for the presence of FB₁ with a median urinary FB₁ concentration of 2.00 and 2.91 ng mg⁻¹ creatinine, respectively. Comparison of the week 8 high and low dose NS treatment groups to placebo revealed a decreased percentage of

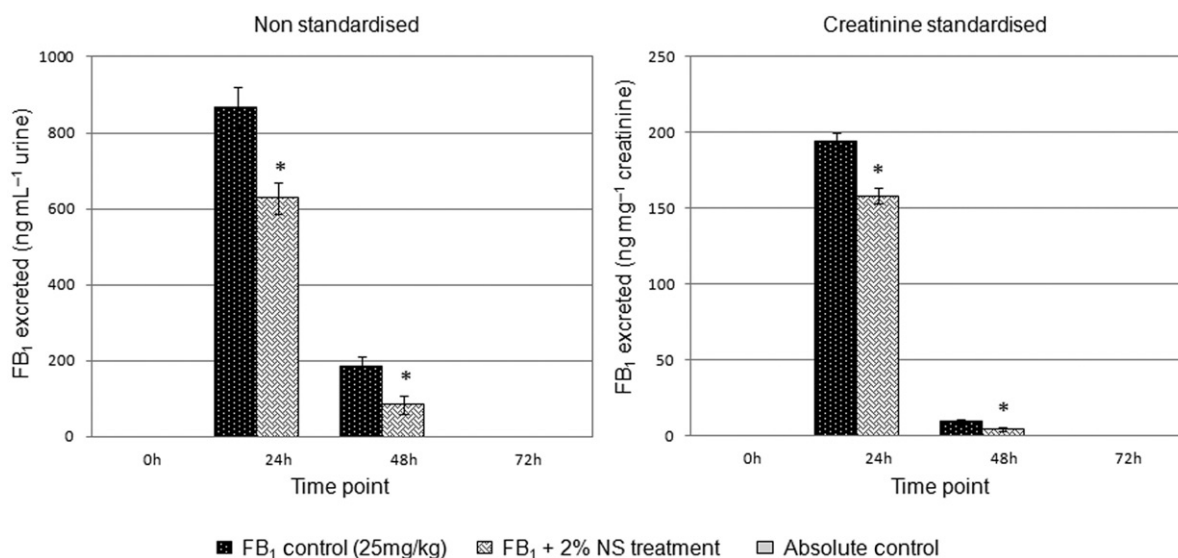


Figure 2. NovaSil reduction of total excreted and creatinine standardised urinary FB₁ in Fischer 344 rats. Data represents the cycle of urinary FB₁ excretion following 25 mg kg⁻¹ dose by gavage. Urinary FB₁ was reduced by 27% in the NS treated group. This reduction was statistically significant ($p \leq 0.05$) when NS treated and non-treated groups were compared using the Kruskal–Wallis and Wilcoxon rank-sum test. Data represents the means and standard error. Non-detectable levels of FB₁ were documented for the control group at all time points.

Table 1. Reduction in FB₁ levels with NovaSil treatment in Fischer 344 rats.

	Absolute control	FB ₁ control	FB ₁ + 2% NS treatment
Urine collection 24 h post-gavage			
% detectable FB ₁ samples	0 ($n=6$)	100 ($n=6$)	100 ($n=6$)
Mean FB ₁ (95% CI)	N.D.	864.68 (719–1010.14)	625.77 (554.17–696.83)
Median FB ₁ (ng ml ⁻¹ urine)	N.D.	892.35	590.41*
Mean FB ₁ (95% CI)	N.D.	194.07 (161.55–226.59)	158.17 (138.21–178.14)
Median FB ₁ (ng mg ⁻¹ creatinine)	N.D.	191.95	154.32*
Urine collection 48 h post-gavage			
% detectable FB ₁ samples	0 ($n=6$)	100 ($n=6$)	100 ($n=6$)
Mean FB ₁ (95% CI)	N.D.	183.07 (110.16–255.98)	83.29 (18.40–148.19)
Median FB ₁ (ng ml ⁻¹ urine)	N.D.	183.65	91.92*
Mean FB ₁ (95% CI)	N.D.	9.38 (6.03–12.72)	4.42 (1.00–7.85)
Median FB ₁ (ng mg ⁻¹ creatinine)	N.D.	9.91	4.95*

Notes: Data represent the mean, confidence interval (CI) and median from each treatment group.

* Significantly different from FB₁ control group, $p \leq 0.05$ (Wilcoxon rank-sum test).

N.D., not detected.

participants with detectable urinary FB₁ (39.1 and 45.5%, respectively). A similar numerical reduction was noted when week 10 high and low dose treatment groups were compared to placebo with 54.3 and 52.5% being positive for FB₁, respectively. Median FB₁ levels for the high dose treatment group were found to be significantly lower than the placebo group at week 8 ($p \leq 0.05$). Median levels for the low dose group were reduced but not found to be significant. Similarly, week 10 median urinary FB₁ levels from both, high and low dose groups were significantly reduced ($p \leq 0.05$) when compared to placebo (Figures 3 and 4; Table 2).

MALDI-TOF analysis

MALDI-TOF mass spectrometry (MTMS) analysis of parent ($M+H^+$), NDA derivatised, and a NDA derivatised + sodium ion ($M+Na^+$) FB₁ confirmed a mass/charge of 723.0, 906.5, and 928.3, respectively (Figure 5A–C). MTMS analysis of FB₁ peaks collected from HPLC analysis of randomly selected positive urine samples verified the presence and identity of FB₁, with peaks at 907.4 ($M+H^+$) and 928.5 ($M+Na^+$) (Figure 5C). The lack of parent peak (~ 722.8) in MTMS analysed samples may provide an indirect indicator of the efficiency of the NDA derivatisation reaction.

Table 2. Reduction in FB₁ levels with NovaSil treatment in the Ghanaian population.

	NovaSil treatment		Control
	High dose (3.0 g day ⁻¹)	Low dose (1.5 g day ⁻¹)	Placebo ^a (1.5 g day ⁻¹)
Week 8 urinary FB₁ analysis			
% detectable FB ₁ samples	39.1 (9/23)	45.5 (10/22)	60.9 (14/23)
Mean FB ₁ (95% CI)	4.05 (0.99–7.10)	5.61 (1.74–9.48)	9.36 (4.51–14.22)
Median FB ₁ (ng ml ⁻¹ urine)	0.19*	0.19	5.45
Mean FB ₁ (95% CI)	1.07 (0.22–1.91)	5.61 (0.81–10.42)	7.12 (1.69–12.52)
Median FB ₁ (ng mg ⁻¹ creatinine)	0.20*	0.20	2.00
Week 10 urinary FB₁ analysis			
% detectable FB ₁ samples	54.3 (19/35)	52.5 (21/40)	72.1 (31/43)
Mean FB ₁ (95% CI)	0.81 (0.53–1.09)	0.73 (0.37–1.08)	5.03 (3.76–6.30)
Median FB ₁ (ng ml ⁻¹ urine)	0.44*	0.21*	5.45
Mean FB ₁ (95% CI)	1.02 (0.45–1.59)	1.69 (0.51–2.87)	6.29 (3.75–8.84)
Median FB ₁ (ng mg ⁻¹ creatinine)	0.28*	0.20*	2.91

Notes: Data represent the mean, confidence interval (CI) and median from each treatment group.

*Significantly different from placebo group, $p \leq 0.05$ (Wilcoxon rank-sum test).

^aPlacebo = microcrystalline cellulose.

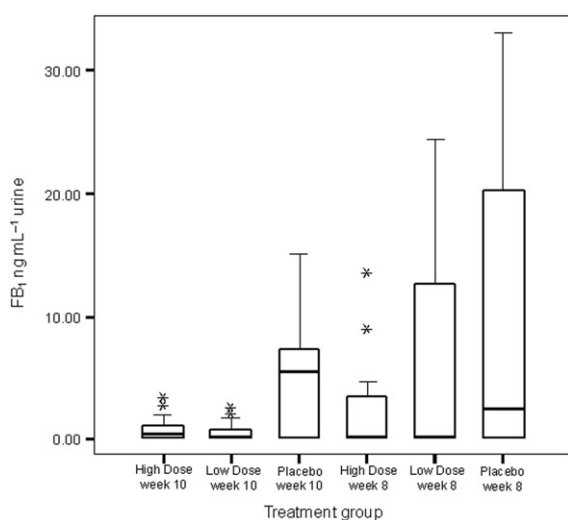


Figure 3. Distribution of urinary FB₁ between treatment groups at weeks 8 and 10. Data represents the distribution of FB₁ (ng FB₁ ml⁻¹ urine) in the various treatment groups based on a NovaSil intervention in Ghana. There was a statistically significant difference among treatments ($p \leq 0.05$). There was a significant difference between the median FB₁ levels in the NS treated groups vs. the placebo group ($p \leq 0.05$). There was not a significant difference between median FB₁ levels between the high and low dose groups ($p = 0.566$).

Discussion

Human exposure to mycotoxins has received increased attention, and recent studies in West Africa have confirmed the co-occurrence of AFs and fumonisins in maize-based foodstuffs found in Cote d'Ivoire and Ghana (Kpodo et al. 2000; Sangare-Tigori et al. 2006).

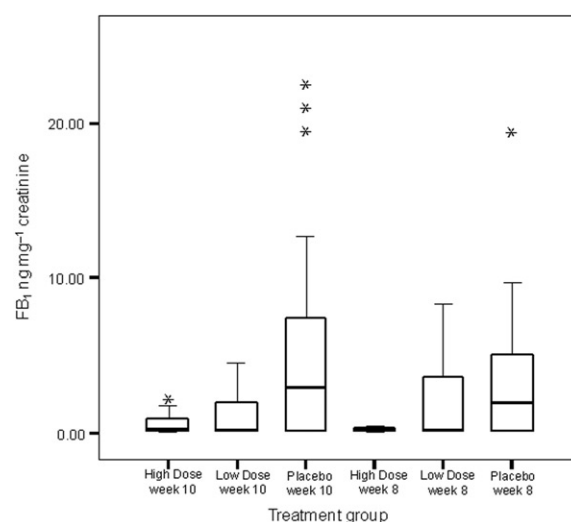


Figure 4. Distribution of creatinine standardised urinary FB₁ between treatment groups at weeks 8 and 10. Data represents the distribution of FB₁ (ng FB₁ mg⁻¹ creatinine) in the various treatment groups based on a NovaSil intervention in Ghana. There was a statistically significant difference among treatments ($p \leq 0.05$). There was a significant difference between the median FB₁ levels in the NS treated groups vs. the placebo group ($p \leq 0.05$). There was not a significant difference between median FB₁ levels in the high and low dose groups ($p = 0.955$).

While surveys have demonstrated that maize-based products are often co-contaminated with diverse mycotoxins; there is a paucity of published data measuring human exposure to fumonisins and AFs. Use of urinary FB₁ as a biomarker of exposure in humans was first proposed by Shetty and Bhat (1998). Turner et al. (1999) reported that analysis of urinary

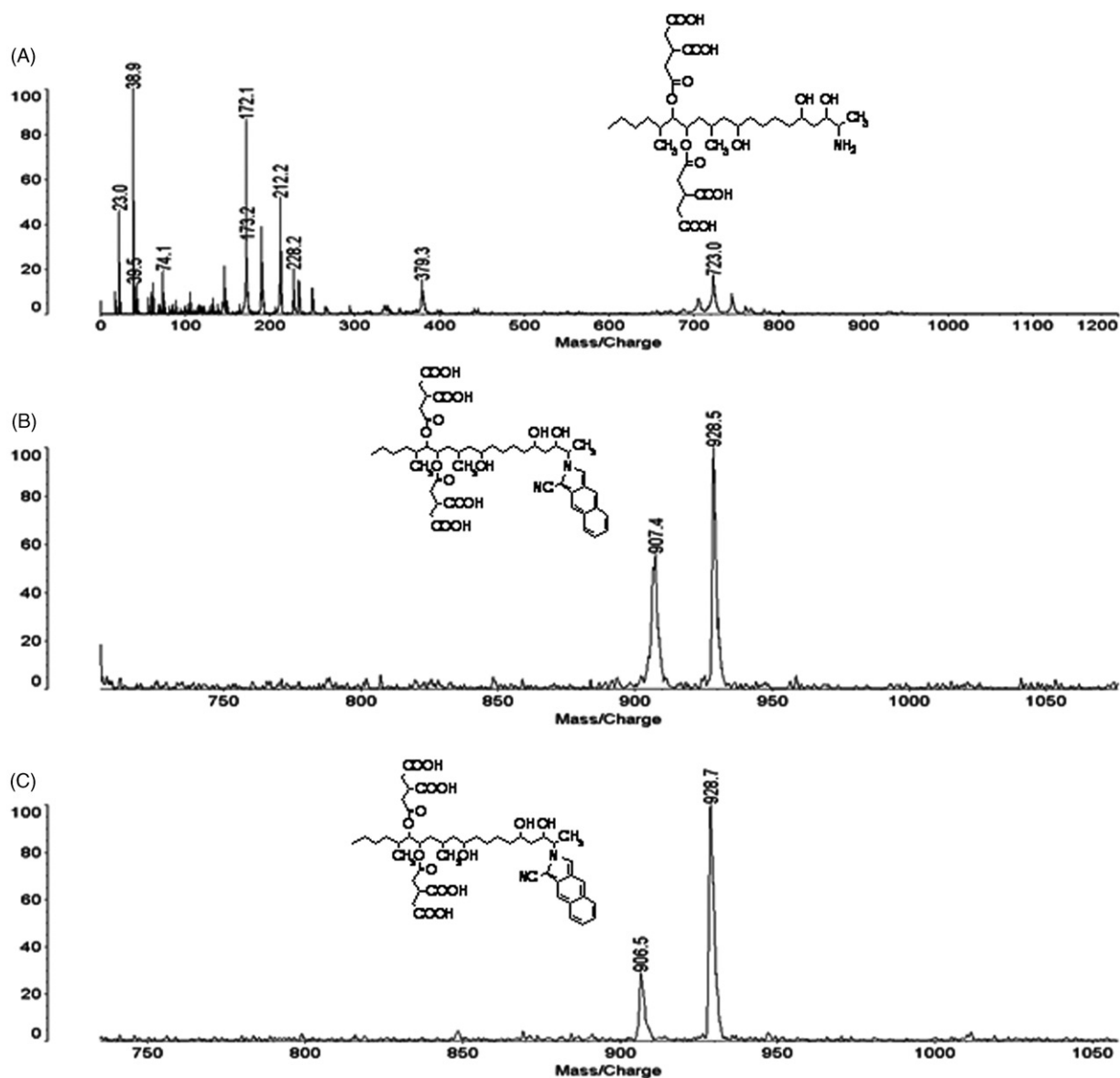


Figure 5. MALDI-TOF mass spectra (MS). MS are based on relative intensity (y-axis) of depicted peaks (highest peak = 100%). (A) MS of underivatized standard FB₁ in water (723 *m/z*). (B) MS of NDA derivatized FB₁ from spiked urine sample (~906.1 *m/z*) and NDA derivative + Na ion (~928 *m/z*). (C) MS of a representative FB₁ positive participant urine sample verifying presence of FB₁.

FB₁ held significant promise as there was a need for a reliable biomarker of exposure to fumonisins. A recent study by Gong et al. (2008) inferred a dose–response relationship between consumption of tortillas containing fumonisins and human urinary FB₁. Data from the present study verified that urinary FB₁ biomarker could be quantified using a rodent model following a single dose exposure. Furthermore, data from our human study in Ghana supports the work by Gong et al. (2008) in Mexico, which suggests that urinary FB₁ is a sensitive biomarker of fumonisin exposure. The decision to use urinary FB₁ as the diagnostic marker of NS efficacy was critical to our *in vivo* analyses. In our rodent study, all animals dosed with FB₁ displayed the urinary FB₁ biomarker within 24 h,

regardless of treatment group. Levels of urinary FB₁ were considerably reduced 48 h after gavage. By 72 h, urinary FB₁ was not detectable. This 48-h window is similar to other urinary biomarkers of mycotoxin exposure (Wang et al. 2008). NS clay has been shown to sorb both aflatoxin and FB₁ *in vitro*, and can decrease the bioavailability and exposure to aflatoxins in humans and animals. Thus, it is possible that NS may also confer protection against FB₁ where dual-contamination of the diet is common. It is then proposed that NS was able to bind FB₁ either at the time of gavage or during the gastrointestinal passage. However, other possibilities may exist. For instance, clays similar to NS have been previously used as mucosa protectants (Droy-Lefaix and Tateo 2006) and

NS may have reduced the urine levels of this mycotoxin by limiting or delaying the contact of FB₁ with the intestinal cell barrier. Our study has two important limitations. One of them is that faecal levels of FB₁ were not measured and this would have been a better verification of our proposed mechanism of action of NS onto FB₁. Another limitation is that our study design does not allow to completely determine if FB₁ was bound to gavage NS or if dietary NS was responsible for the reduction of FB₁ in urine. Nonetheless, in our rodent model, NovaSil (either by gavage or dietary) was able to significantly reduce urinary FB₁ by 20% at 24 h and by 50% at 48 h post-gavage. These data suggest that NS may be a suitable enterosorbent for the reduction of fumonisin exposure, confirming previous reports in our laboratory (Lemke 2000).

In our Ghanaian study, 60.9 and 72.1% of analysed samples from the placebo group contained detectable levels of urinary FB₁, compared to 75% detected in a Mexican population consuming a high intake of maize-based tortillas (Gong et al. 2008). The range of FB₁ in urine samples from our study varied from non-detectable to 33.12 ng FB₁ ml⁻¹ urine, higher than values reported by Gong et al. (2008) (non-detectable to 9.31 ng FB₁ ml⁻¹ urine). The week 8 and 10 mean FB₁ levels measured in participants of the placebo group in our study (9.36 and 5.03 ng FB₁ ml⁻¹ urine, respectively) were higher than mean urinary FB₁ concentration reported in Mexico (0.147 ng FB₁ ml⁻¹ urine) (Gong et al. 2008). Based on ~1% urinary excretion, an estimated metabolic rate of 1500 ml urine day⁻¹ and the average size of an adult (70 kg), it can be estimated that the exposure to FB₁ at week 8 was 20.06 µg kg⁻¹ bw day⁻¹ and 10.78 µg kg⁻¹ bw day⁻¹ at week 10 in our placebo groups (Shephard et al. 1994; Fodor et al. 2008). More importantly, recent studies in a South African population consuming corn naturally contaminated with FBs have estimated an average of 0.075% urinary excretion rate of FB₁ (Van der Westhuizen et al. 2011). If this is the case, our Ghanaian population, may have been exposed to 10 times more than what we have previously estimated. These values are higher than the FB₁ levels that could be extrapolated from the Mexican study (0.368 µg kg⁻¹ bw day⁻¹) (Gong et al. 2008). These differences between Mexican and Ghanaian population biomarker levels could be attributed to variations in maize storage, processing, and/or consumption. A market survey in Ghana detected levels of FB₁ between 70 and 2621 (mean 608.8) µg kg⁻¹ in maize intended for human consumption (Jolly et al. 2006). Based on an average maize intake of 369.5 g day⁻¹ (MacIntyre et al. 2002), a 1% carryover of FB₁ from the diet, and an average urinary excretion (1500 ml), Ghanaians from our site could potentially excrete urinary FB₁ levels between 0.172 to 6.46 ng ml⁻¹ urine. Actual mean

biomarker levels measured in the placebo group of the present study were similar to this estimate, with measured values of 9.36 and 5.03 ng FB₁ ml⁻¹ urine in the week 8 and week 10 groups, respectively.

In the previous clinical trial in Ghana, significant decreases in median urinary aflatoxin M₁ levels were observed at 12 weeks after NS intervention ($p = 0.0445$) (Wang et al. 2008). Since samples that were collected at 12 weeks were depleted for aflatoxin biomarker analysis, we utilised samples at weeks 8 and 10, to assess the effect of NS treatment on urinary FB₁ levels. With NS, the week 8 samples had mean values of 4.05 and 5.61 ng ml⁻¹ urine in the high and low dose treatment groups, respectively, compared with 9.36 ng ml⁻¹ in the placebo group. At week 10, samples had lower mean values of 0.81 and 0.73 ng FB₁ ml⁻¹ urine in the high and low dose treatment groups, respectively, versus 5.03 ng FB₁ ml⁻¹ urine in the placebo group. While the lower levels of FB₁ measured in the participants of low dose and high dose NS are consistent with the effects of NS in reduction of FB₁ bioavailability, the possibility that the groups (placebo, low dose NS and high dose NS) were consuming different levels of FB₁ in the diets, represents a valid question. Nonetheless, the diets ingested by participants were likely to be uniform throughout the study period since the sources of maize tend to be constrained community wide. Furthermore, the effect of NS or placebo on FB₁ urinary levels was observed at two different time points.

When evaluating the means between the placebo (humans) or FB₁ control group (rats) and NS treated groups the reduction in urinary biomarker levels due to NS was more evident in the human study (a 49% reduction in week 8 and an 85% reduction in week 10) than in the rat study (a 19% reduction after 24 h and a 50% reduction after 48 h). This disparity between the mean reductions in FB₁ excretion (rats vs. humans) is not unexpected. In the clinical trial, Ghanaians were chronically exposed to FB₁ through the diet and treatment with NS (by capsules) occurred for a period of 3 months, whereas, in the rat study, group C was given FB₁ as a onetime dose and NS treatment only lasted for 2 weeks. Additionally, it is known that gastrointestinal transit times vary according to size and weight of different species. Thus, it is possible that, in the humans, NS and FB₁ had a longer time to interact resulting in a larger reduction in FB₁ excretion. Nevertheless, in the rat and human studies, NS significantly reduced the excretion of FB₁ biomarker in the urine. Further studies are warranted to establish whether a time-related effect exists allowing NS treatment to exert more effect at later time points. Based on the 10-week mean urine values in the NS treated groups, exposure could be estimated at 1.72 and 1.54 µg kg⁻¹ bw day⁻¹ for the high and low dose participants, respectively. Importantly, following consumption with NS, participant exposure to FB₁ was

reduced below the JECFA provisional maximum tolerable daily intake for FB₁ of 2 µg kg⁻¹ bw day⁻¹ (JECFA 2001). Based on our previous research, NS clay has displayed: (1) favourable thermodynamic characteristics of aflatoxin sorption; (2) tolerable levels of priority metals, dioxins/furans and other hazardous contaminants; (3) safety and efficacy in multiple animal species and humans; (4) safety and efficacy in long-term animal studies; and (5) negligible interactions with vitamins, iron and zinc and other micronutrients. Additionally, recent work in our laboratory has shown that non-framework micronutrients associated with NS (i.e. Sr²⁺) are bioavailable suggesting that NS has the potential to act as a supplement for these micronutrients as well as a mycotoxin enterosorbent in humans and animals (Afriyie-Gyawu et al. 2008). Further work is warranted to delineate the specific molecular mechanism(s) and surface chemistry involved in the sorption process of FB₁ onto NS and to confirm the efficacy and safety of NS clay as a multifunctional intervention for aflatoxins and fumonisins in animals and humans.

Mycotoxins have been linked to death and disease, with an estimated 4.5 billion people facing uncontrolled exposure in developing countries (Williams et al. 2004). The mycotoxin problem to public health is longstanding, unavoidable, and seemingly inextricable. Frequent and concurrent exposure to aflatoxins and fumonisins may enhance/synergise the toxic, carcinogenic, and teratogenic effects of these foodborne contaminants. This study is the first to demonstrate the efficacy of an enterosorbent intervention for FB₁ exposure using fumonisin biomarkers in human urine. Importantly, it also illustrates that a susceptible population in Ghana co-exposed to aflatoxins and fumonisins may be protected from the adverse effects of these mycotoxins using NS clay. As an intervention, capsules or other dose forms of NS could be used in clinical practice for "acute emergencies" and dietary practice for "chronic exposures." Additional studies are planned to confirm these findings in long-term clinical intervention trials in Ghana and to look at the nutritional implications of NS supplementation of the diet. Clay-based enterosorbent strategies represent a novel approach to the problem in populations that are at high risk for mycotoxicosis. A similar clinical intervention with NS in a population at risk for hepatocellular carcinoma (HCC) in the USA is ongoing, and will examine dietary and environmental risk factors for HCC including aflatoxins and fumonisins and the effect of NS intervention.

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References

- Abnet CC, Borkowf CB, Qiao YL, Albert PS, Wang E, Merrill Jr. AH, Mark SD, Dong ZW, Taylor PR, Dawsey SM. 2001. Sphingolipids as biomarkers of fumonisin exposure and risk of oesophageal squamous cell carcinoma in china. *Cancer Causes Control*. 12: 821–828.
- Afriyie-Gyawu E, Mackie J, Dash B, Wiles M, Taylor J, Huebner H, Tang L, Guan H, Wang J-S, Phillips TD. 2005. Chronic toxicological evaluation of dietary NovaSil Clay in Sprague-Dawley rats. *Food Addit Contam*. 22:259–269.
- Afriyie-Gyawu E, Ankrah N-A, Huebner HJ, Ofosuhene M, Kumi J, Johnson NM, Tang L, Xu L, Jolly PE, Ellis WO, et al. 2008. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis. I. Study design and clinical outcomes. *Food Addit Contam A*. 25:622–634.
- Cai Q, Tang L, Wang JS. 2007. Validation of fumonisin biomarkers in F344 rats. *Toxicol Appl Pharmacol*. 225:28–39.
- Carlson DB, Williams DE, Spitsbergen JM, Ross PF, Bacon CW, Meredith FI, Riley RT. 2001. Fumonisin B1 promotes aflatoxin B1 and *N*-methyl-*N'*-nitro-nitrosoguanidine-initiated liver tumors in rainbow trout. *Toxicol Appl Pharmacol*. 172:29–36.
- Cho YH, Yoo HS, Min JK, Lee EY, Hong SP, Chung YB, Lee YM. 2002. Comparative study of naphthalene 2,3-dicarboxaldehyde and *o*-phthalaldehyde fluorogenic reagents for chromatographic detection of sphingoid bases. *J Chromatogr A*. 977:69–76.
- Chu FS, Li GY. 1994. Simultaneous occurrence of fumonisin B1 and other mycotoxins in moldy corn collected from the People's Republic of China in regions with high incidences of oesophageal cancer. *Appl Environ Microbiol*. 60:847–52.
- Droy-Lefaix MT, Tateo F. 2006. Handbook of clay science. Kidlington (UK): Elsevier. Chapter 11.6, Clays and clay minerals as drugs; p. 744.
- Fodor J, Balogh K, Weber M, Mezes M, Kametler L, Posa R, Mamet R, Bauer J, Horn P, Kovacs F, et al. 2008. Absorption, distribution and elimination of fumonisin B1 metabolites in weaned piglets. *Food Addit Contam A*. 25:88–96.
- Gong YY, Torres-Sanchez L, Lopez-Carrillo L, Peng JH, Sutcliffe AE, White KL, Humpf HU, Turner PC, Wild CP. 2008. Association between tortilla consumption and human urinary fumonisin B1 levels in a Mexican population. *Cancer Epidemiol Biomarkers Prev*. 17:688–694.
- He X, Dagan A, Gatt S, Schuchman EH. 2005. Simultaneous quantitative analysis of ceramide and sphingosine in mouse blood by naphthalene 2,3 dicarboxaldehyde derivatisation after hydrolysis with ceramidase. *Anal Biochem*. 340:113–122.

- He Q, Suzuki H, Sharma N, Sharma RP. 2006. Ceramide synthase inhibition by fumonisin B1 treatment activates sphingolipid-metabolizing systems in mouse liver. *J Toxicol Sci.* 94:388–397.
- IARC. 2002. Monographs on the evaluation of carcinogenic risks to humans; some traditional herbal medicines, some mycotoxins, naphthalene and styrene. Vol. 82. Lyon: IARC.
- JECFA 2001. Evaluation of certain mycotoxins in food. Fifty-sixth report of the Joint FAO/WHO Expert Committee on Food Additives. WHO technical report series 906:1–62; Geneva, Switzerland: WHO.
- Jiang Y, Jolly PE, Ellis WO, Wang JS, Phillips TD, Williams JH. 2005. Aflatoxin B1 albumin adduct levels and cellular immune status in Ghanaians. *Int Immunol.* 17:807–814.
- Jolly PE, Jiang Y, Ellis W, Awuah R, Nnedu O, Phillips T, Wang JS, Afriyie-Gyawu E, Tang L, Person S, et al. 2006. Determinants of aflatoxin levels in Ghanaians: socio-demographic factors, knowledge of aflatoxin and food handling and consumption practices. *Int J Hyg Environ Health.* 209:345–358.
- Kpodo K, Thrane U, Hald B. 2000. Fusaria and fumonisins in maize from Ghana and their co-occurrence with aflatoxins. *Int J Food Microbiol.* 61:147–157.
- Lemke SL. 2000. Investigation of clay based strategies for the protection of animals from the toxic effects of selected mycotoxins [PhD dissertation]. [College Station (TX)]: Texas A&M University.
- MacIntyre UE, Kruger HS, Venter CS, Vorster HH. 2002. Dietary intakes of an African population in different stages of transition in the North West Province South Africa: the THUSA study. *Nutr Res.* 22:239–256.
- Marasas WF, Riley RT, Hendricks KA, Stevens VL, Sadler TW, Gelineau-van Waes J, Missmer SA, Cabrera J, Torres O, Gelderblom WC, et al. 2004. Fumonisin disrupt sphingolipid metabolism, folate transport, and neural tube development in embryo culture and in vivo: a potential risk factor for human neural tube defects among populations consuming fumonisin-contaminated maize. *J Nutr.* 134:711–716.
- McKean C, Tang L, Tang M, Billam M, Wang Z, Theodorakis CW, Keandall RJ, Wang J-S. 2006. Comparative acute and combinative toxicity of aflatoxin B1 and fumonisin B1 in animals and human cells. *Food Chem Toxicol.* 44:868–876.
- Neupert S, Russell WK, Russell DH, López Jr. JD, Predel R, Nachman RJ. 2009. Neuropeptides in Heteroptera: identification of allatotropin-related peptide and tachykinin-related peptides using MALDI-TOF mass spectrometry. *Peptides.* 30:483–488.
- Phillips TD, Afriyie-Gyawu E, Williams J, Huebner H, Ankrah NA, Ofori-Adjei D, Jolly P, Johnson N, Taylor J, Marroquin-Cardona A, et al. 2008. Reducing human exposure to aflatoxin through the use of clay: a review. *Food Addit Contam A.* 25:134–145.
- Sabourdy F, Kedjouar B, Sorli SC, Colié S, Milhas D, Salma Y, Levade T. 2008. Functions of sphingolipid metabolism in mammals—lessons from genetic defects. *Biochim Biophys Acta.* 1781:145–183.
- Sangare-Tigori B, Moukha S, Kouadio HJ, Betbeder A-M, Dano DS, Creppy ED. 2006. Co-occurrence of aflatoxin B1, fumonisin B1, ochratoxin A and zearalenone in cereals and peanuts from Cote d'Ivoire. *Food Addit Contam.* 23:1000–1007.
- Shephard GS, Thiel PG, Stockenstrom S, Sydenham EW. 1996. Worldwide survey of fumonisin contamination of corn and corn-based products. *J Assoc Off Anal Chem Int.* 79:671–687.
- Shephard GS, Thiel PG, Sydenham EW, Alberts JF, Cawood ME. 1994. Distribution and excretion of a single dose of the mycotoxin fumonisin B1 in a non-human primate. *Toxicol.* 32:735–741.
- Shephard GS, Marasas WF, Burger HM, Somdya NI, Rheeder JR, Van Der Westhuizen L, Gatyeni P, Van Schalkwyk DJ. 2007. Exposure assessment for fumonisins in the former Transkei region of South Africa. *Food Addit Contam.* 24:621–629.
- Shetty PH, Bhat RV. 1998. Sensitive method for the detection of fumonisin B1 in human urine. *J Chromatogr B.* 705:171–173.
- Solfizzo M, Chulze SN, Mallmann C, Visconti A, De Girolamo A, Rojo F, Torres A. 2004. Comparison of urinary sphingolipids in human populations with high and low maize consumption as a possible biomarker of fumonisin dietary exposure. *Food Addit Contam.* 21:1090–1095.
- Stockmann-Juvala H, Savolainen K. 2008. A review of the toxic effects and mechanisms of action of fumonisin B1. *Hum Exp Toxicol.* 27:799–809.
- Turner PC, Nikiema P, Wild CP. 1999. Fumonisin contamination of food: progress in development of biomarkers to better assess human health risks. *Mutat Res.* 443:81–93.
- Ueno Y, Iijima K, Wang SD, Sugiura Y, Sekijima M, Tanaka T, Chen C, Yu SZ. 1997. Fumonisin as a possible contributory risk factor for primary liver cancer: a 3-year study of corn harvested in Haimen, China, by HPLC and ELISA. *Food Chem Toxicol.* 35:1143–1150.
- Van der Westhuizen L, Shephard GS, Burger HM, Rheeder JP, Gelderblom WCA, Wild CP, Gong YY. 2011. Fumonisin B1 as a urinary biomarker of exposure in a maize intervention study among South African subsistence farmers. *Cancer Epidemiol. Biomarkers Prev.* 20:483–489.
- Voss KA, Howard PC, Riley RT, Sharma RP, Bucci TJ, Lorentzen RJ. 2002. Carcinogenicity and mechanism of action of fumonisin B1: a mycotoxin produced by *Fusarium moniliforme* (= *F. verticillioides*). *Cancer Detect Prev.* 26:1–9.
- Wang P, Afriyie-Gyawu E, Tang Y, Johnson NM, Xu L, Tang L, Huebner HJ, Ankrah NA, Ofori-Adjei D, Ellis W, et al. 2008. NovaSil clay intervention in Ghanaians at high risk for aflatoxicosis II. Reduction in biomarkers of aflatoxin exposure in blood and urine. *Food Addit Contam A.* 25:622–634.
- Williams JH, Phillips TD, Jolly PE, Stiles JK, Jolly CM, Aggarwal D. 2004. Human aflatoxicosis in developing countries: a review of toxicology, exposure, potential health consequences, and interventions. *Am J Clin Nutr.* 80:1106–1122.
- Wogan GN. 1992. Aflatoxins as risk factors for hepatocellular carcinoma in humans. *Cancer Res.* 52:2114s–2118s.